# PET studies of cerebral metabolism in Parkinson Disease

William J. Powers

Published online: 11 November 2009 © Springer Science + Business Media, LLC 2009

Abstract A defect in cerebral energy production due to dysfunction of the mitochondrial electron transport system (ETS) has been postulated to be important in the pathogenesis of Parkinson Disease (PD). However, direct in vivo measurements of cerebral mitochondrial function are scant and inconsistent. We directly investigated cerebral mitochondrial function in vivo with positron emission tomography (PET) in 12 patients with early, never-medicated PD and 12 age-matched normal controls by combined measurements of the cerebral metabolic rate of oxygen (CMRO<sub>2</sub>) and the cerebral metabolic rate of glucose (CMRglc). Instead of the decrease in CMRO<sub>2</sub> and CMRO<sub>2</sub>/CMRglc molar ratio characteristic of defects in mitochondrial oxidative metabolism, there was a statistically significant 24% general increase in CMRO2 and no change in CMRO<sub>2</sub>/CMRglc. Since PD symptoms were already manifest, reduced oxidative activity of the mitochondrial ETS cannot be a primary mechanism of neuronal death in early PD. This increase in metabolism could reflect the increased energy requirements of an injured brain or an uncoupling of ATP production from oxidation in the terminal stage of oxidative phosphorylation. Which is the case in early PD and whether these metabolic abnormalities are important in the pathogenesis of PD will require further study.

**Keywords** Parkinson Disease · Cerebral oxygen metabolism · Cerebral glucose metabolism · Mitochondria

# Introduction

Dysfunction of mitochondrial oxidative metabolism has been implicated in the pathogenesis of Parkinson Disease (PD) (Abou-Sleiman et al. 2006). Diminished activity in complex I of the mitochondrial electron transport system (ETS) in post-mortem brain tissue has been reported in cortex and substantia nigra (Schapira 1994; Keeney et al. 2006). However, in vivo evidence of cerebral mitochondrial electron transport dysfunction is scant and inconsistent. Low ATP levels have been reported in the cortex in two studies (Piert et al. 1996; Rango et al. 2006). Overall cerebral oxygen metabolism has been reported to be not different from normal, but with relative increases in the basal ganglia opposite to the most symptomatic side (Wolfson et al. 1985; Leenders et al. 1985). These in vivo studies were performed in patients with mean disease duration of 6-7 years that was treated with L-dopa. If a defect in mitochondrial electron transport is important in the pathogenesis of PD, it will be present early in the course of the disease and prior to the possibly confounding effects of drug therapy.

Specific defects in mitochondrial ETS decrease CMRO<sub>2</sub> proportionately more than CMRglc (fewer moles of oxygen consumed per mole of glucose metabolized), thereby producing a reduction in the CMRO<sub>2</sub>/CMRglc molar ratio below the normal value of 5.6 (Brierley et al. 1977; Frackowiak et al. 1988). Therefore, in vivo assessment of mitochondrial energy metabolism requires combined measurement of CMRO<sub>2</sub> and CMRglc, which had not previously performed in Parkinson Disease.

## PET studies of early PD at Washington University

We directly measured CMRO<sub>2</sub> and CMRglc in vivo with positron emission tomography (PET) in 12 early, never-

W. J. Powers (🖂)

Department of Neurology, School of Medicine, University of North Carolina at Chapel Hill, 170 Manning Drive, Rm 2131, CB #7025, Chapel Hill, NC 27599-7025, USA e-mail: powersw@neurology.unc.edu

Table 1Bilateral cerebral andsubstantia nigra metabolism inearly Parkinson Disease (fromPowers et al. 2008)

CMRO<sub>2</sub>—cerebral metabolic rate of oxygen (micromoles  $100 \text{ g}^{-1} \text{ min}^{-1}$ ), CMRglc—cerebral metabolic rate of glucose (micromoles  $100 \text{ g}^{-1} \text{ min}^{-1}$ ), Values are mean  $\pm$  SD

<sup>a</sup> Primary analysis

	Number	CMRO <sub>2</sub>	CMRglc	CMRO <sub>2</sub> /CMRglc
Bihemispheric				
Normal controls	12	$115 \pm 25$	$20.7 \pm 2.6$	5.6±1.3
Parkinson Disease	12	$143 \pm 36$	$23.8 \pm 4.4$	$6.15 \pm 1.6$
<i>t</i> -test		$p = .037^{a}$	<i>p</i> =.056	<i>p</i> =.39
Substantia nigra				
Normal controls	10	$110 \pm 71$	$15.9 \pm 2.3$	$6.78 \pm 3.91$
Parkinson Disease	10	$147 \pm 91$	$17.9 \pm 3.2$	$8.36{\pm}4.91$
<i>t</i> -test		<i>p</i> =.313	<i>p</i> =.122	<i>p</i> =.437

medicated participants with PD and compared to 12 agematched normal controls. All participants also underwent magnetic resonance scanning to permit correction of the PET data for brain atrophy and to permit accurate localization of substantia nigra, putamen and globus pallidus for regional measurements (Powers et al. 2008).

The 12 participants (6 men/6 women) with PD were 44– 77 years old (mean 60). Symptoms had been present for 8– 48 months (mean 22). Hoehn and Yahr stages were: 2 Stage 1, 1 Stage 1.5, and 9 Stage 2 (Hoehn and Yahr 1967). Twelve normal control subjects from a cohort of 23 were matched by age to the 12 participants with PD without reference to any PET data. Their ages were 45–71 (mean 61) years. There were 6 men and 6 women.

There was a statistically significant 24% increase in hemispheric CMRO<sub>2</sub> in PD (p=.037) (Table 1). This change was in the opposite direction of the decrease that occurs with defects in mitochondrial electron transport. Hemispheric CMRglc also was increased by 15% and CMRO<sub>2</sub>/CMRglc was increased by 10%. Both of these changes are also in the opposite direction that occurs with defects in mitochondrial electron transport. Examination of the confidence intervals for the differences between the two groups for these latter two measurements demonstrates that there is less than a 6% chance that CMRglc is lower in PD

by any amount and only a 10% chance that  $\rm CMRO_2/$  CMRglc is reduced by 10% or more.

Similar results, albeit with more measurement imprecision as expected for regional data, were found in the substantia nigra, putamen and globus pallidus (Tables 1 and 2). Analysis of regional/hemispheric ratios for putamen and globus pallidus showed no difference between controls and participants with PD indicating that the increases in regional metabolism were primarily a reflection of overall brain changes.

### Previous PET studies of cerebral metabolism in PD

Previous PET studies of cerebral metabolism in PD have yielded mixed results. In five studies of global CMRglc, four have reported reductions of approximately 20% and one reported no significant difference compared to agematched controls (Kuhl et al. 1984; Leenders et al. 1985; Eidelberg et al. 1993, 1994; Piert et al. 1996). In one of these studies, reductions in global CMRglc were seen only after L-dopa was administered suggesting that the reduction in metabolism may be at least in part due to medication effects (Berding et al. 2001). Berding et al. (2001) have suggested that hypometabolism parallels disease duration.

 Table 2
 Bilateral basal ganglia metabolism in early Parkinson Disease (from Powers et al. 2008)

	Number	CMRO <sub>2</sub>	Regional/Hemispheric CMRO <sub>2</sub> ratio	CMRglc	Regional/Hemispheric CMRglc ratio
Putamen					
Normal controls	12	138±27	$1.21 \pm 0.06$	24.2±4.1	$1.17{\pm}0.13$
Parkinson Disease	12	$175 \pm 40$	$1.23 \pm 0.19$	29.8±5.0	$1.26 \pm 0.11$
<i>t</i> -test		<i>p</i> =.016	<i>p</i> =.86	<i>p</i> =.007	p=.08
Globus pallidus					
Normal controls	12	$110 \pm 39$	$0.95 {\pm} 0.19$	$16.8 {\pm} 2.8$	$0.81 {\pm} 0.07$
Parkinson Disease	12	137±37	$0.98 {\pm} 0.23$	$20.6 \pm 2.8$	$0.88 \pm 0.12$
<i>t</i> -test		<i>p</i> =.097	<i>p</i> =.702	<i>p</i> =.003	<i>p</i> =.093

 $CMRO_2$ —cerebral metabolic rate of oxygen (micromoles 100 g<sup>-1</sup> min<sup>-1</sup>), CMRglc—cerebral metabolic rate of glucose (micromoles 100 g<sup>-1</sup> min<sup>-1</sup>), Values are mean  $\pm$  SD

Thus, these reported changes in CMRglc likely reflect a consequence of the PD disease process. We deliberately chose to study patients with very early disease to try to determine if there was metabolic dysfunction that caused PD. The mean disease duration of 22 months in our study was substantially shorter than in these previous studies where it ranged from 4 to 15 years. Our analysis using absolute and relative measurements showing a trend toward increased global CMRglc in very early PD supports the theory that the reported reductions in metabolism are a consequence, not a cause, of the disease.

Regional basal ganglia metabolism in PD measured with PET has been reported to be increased, decreased or unchanged (Kuhl et al. 1984; Rougemont et al. 1984; Martin et al. 1984; Wolfson et al. 1985; Leenders et al. 1985; Mohr et al. 1992; Eidelberg et al. 1993, 1994, 1995; Piert et al. 1996). In these studies, findings are dependent on whether analysis is performed using absolute values, relative values or more sophisticated image analysis techniques such as statistical parametric mapping or scaled subprofile modeling (Eidelberg et al. 1993, 1995; Piert et al. 1996). The validity of analysis methods that use normalization based on whole brain values for assessing relative basal ganglia metabolism has recently been challenged (Borghammer et al. 2009). We found no evidence for relative increases or decreases in basal ganglia CMRO<sub>2</sub> or CMRglc in early PD using either absolute values or normalization based on whole brain values.

#### Increased cerebral oxidative metabolism in early PD

The pathophysiological basis for the increase in cerebral oxidative metabolism in early PD is not known. Based on the classic study by Wooten and Collins (1981) who described transient glucose hypermetabolism focally restricted to ipsilateral globus pallidus following unilateral 6hydroxydopamine lesions of the substantia nigra in rats, increased metabolism in basal ganglia structures has been ascribed to loss of dopaminergic inhibitory pathways (Martin et al. 1984; Wolfson et al. 1985; Eidelberg et al. 1993). A generalized loss of inhibitory dopaminergic input throughout the brain is a possible cause for the general increase in metabolism. Alternatively, this general increase in CMRO<sub>2</sub> could be due not to increased metabolic demand, but to an uncoupling of ATP production from oxidation in the terminal stage of oxidative phosphorylation. Uncoupling (dysfunction of Complex V ATP synthase) produces an increase in both CMRO<sub>2</sub> and CMRglc similar to what we observed (Patel and Brewer 2003; Tretter and Adam-Vizi 2007). Whether uncoupling of oxidative phosphorylation occurs in early PD and whether or not it is important in the pathogenesis of PD will require further study.

#### Summary and conclusions

In summary, we found a generalized increase in cerebral oxygen metabolism in never-medicated patients with early PD. Since PD symptoms were already manifest, we can exclude reduced oxidative activity of the mitochondrial ETS as a pathogenic mechanism of their disease. Thus, while defects in mitochondrial ETS may be present in some patients with PD, the absence of defects in cerebral oxidative metabolism in these12 patients with early PD indicates that dysfunction of ETS-mediated oxidation cannot be essential to the pathogenesis of neuronal death in early PD.

Acknowledgments This research was supported by USPHS grants NS 41771 and NS35966, the Lillian Strauss Institute for Neuroscience and the Elliot Stein Family Fund of the Barnes-Jewish Hospital Foundation, the Huntington's Disease Society of America Center of Excellence at Washington University, the American Parkinson Disease Association (APDA) Advanced Center for Research at Washington University, and the Greater St. Louis Chapter of the APDA and the H. Houston Merritt Professorship of Neurology at the University of North Carolina at Chapel Hill. I would like to thank my colleagues Tom Videen, Joanne Markham, Kevin Black, Nima Golchin, Joel Perlmutter, Lennis Lich, John Hood, Lori-McGee-Minnich, Susanne Fritsch and the Washington University Cyclotron Staff for their assistance.

#### References

- Abou-Sleiman PM, Muqit MM, Wood NW (2006) Expanding insights of mitochondrial dysfunction in Parkinson's disease. Nat Rev Neurosci 7:207–219
- Berding G, Odin P, Brooks DJ, Nikkhah G, Matthies C, Peschel T, Shing M, Kolbe H, van Den Hoff J, Fricke H, Dengler R, Samii M, Knapp WH (2001) Resting regional cerebral glucose metabolism in advanced Parkinson's disease studied in the off and on conditions. Mov Disord 16:1014–1022
- Borghammer P, Cumming P, Aanerud J, Förster S, Gjedde A (2009) Subcortical elevation of metabolism in Parkinson's disease—a critical reappraisal in the context of global mean normalization. Neuroimage 47:1514–1521
- Brierley JB, Prior PF, Calverley J, Brown AW (1977) Cyanide intoxication in Macaca mulatta. Physiological and neuropathological aspects. J Neurol Sci 31:133–157
- Eidelberg D, Takikawa S, Moeller JR, Dhawan V, Redington K, Chaly T, Robeson W, Dahl JR, Margouleff D, Fazzini E (1993) Striatal hypometabolism distinguishes striatonigral degeneration from Parkinson's disease. Ann Neurol 33:518–527
- Eidelberg D, Moeller JR, Dhawan V, Spetsieris P, Takikawa S, Ishikawa T, Chaly T, Robeson W, Margouleff D, Przedborski S (1994) The metabolic topography of Parkinsonism. J Cereb Blood Flow Metab 14:783–801
- Eidelberg D, Moeller JR, Ishikawa T, Dhawan V, Spetsieris P, Chaly T, Belakhlef A, Mandel F, Przedborski S, Fahn S (1995) Early differential diagnosis of Parkinson's disease with 18Ffluorodeoxyglucose and positron emission tomography. Neurology 45:1995–2004
- Frackowiak RSJ, Herold S, Petty RK, Morgan-Hughes JA (1988) The cerebral metabolism of glucose of oxygen measured with positron tomography in patients with mitochrondrial diseases. Brain 111:1009–1024

- Hoehn MM, Yahr MD (1967) Parkinsonism: onset, progression and mortality. Neurology 17:427–442
- Keeney PM, Xie J, Capaldi RA, Bennett JP Jr (2006) Parkinson's disease brain mitochondrial complex I has oxidatively damaged subunits and is functionally impaired and misassembled. J Neurosci 26:5256–5264
- Kuhl DE, Metter EJ, Riege WH (1984) Patterns of local cerebral glucose utilization determined in Parkinson's disease by the [18F]fluorodeoxyglucose method. Ann Neurol 15:419–424
- Leenders KL, Wolfson L, Gibbs JM, Wise RJ, Causon R, Jones T, Legg NJ (1985) The effects of L-DOPA on regional cerebral blood flow and oxygen metabolism in patients with Parkinson's disease. Brain 108:171–191
- Martin WR, Beckman JH, Calne DB, Adam MJ, Harrop R, Rogers JG, Ruth TJ, Sayre CI, Pate BD (1984) Cerebral glucose metabolism in Parkinson's disease. Can J Neurol Sci 11:169–173
- Mohr E, Mann UM, Miletich RS, Sampson M, Goldberg TE, Grimes JD, Chase TN (1992) Neuropsychological and glucose metabolic profiles in asymmetric Parkinson's disease. Can J Neurol Sci 19:163–169
- Patel JR, Brewer GJ (2003) Age-related changes in neuronal glucose uptake in response to glutamate and beta-amyloid. J Neurosci Res 72:527–536

- Piert M, Koeppe RA, Giordani B, Minoshima S, Kuhl DE (1996) Determination of regional rate constants from dynamic FDG-PET studies in Parkinson's disease. J Nucl Med 37:1115–1122
- Powers WJ, Videen TO, Markham J, Black KJ, Golchin N, Perlmutter JS (2008) Cerebral mitochondrial metabolism in early Parkinson's disease. J Cereb Blood Flow Metab 28:1754–1760
- Rango M, Bonifati C, Bresolin N (2006) Parkinson's disease and brain mitochondrial dysfunction: a functional phosphorus magnetic resonance spectroscopy study. J Cereb Blood Flow Metab 26:283–290
- Rougemont D, Baron JC, Collard P, Bustany P, Comar D, Agid Y (1984) Local cerebral glucose utilisation in treated and untreated patients with Parkinson's disease. J Neurol Neurosurg Psychiatry 47:824–830
- Schapira AH (1994) Evidence for mitochondrial dysfunction in Parkinson's disease—a critical appraisal. Mov Disord 9:125–138
- Tretter L, Adam-Vizi V (2007) Uncoupling is without an effect on the production of reactive oxygen species by in situ synaptic mitochondria. J Neurochem 103:1864–1871
- Wolfson LI, Leenders KL, Brown LL, Jones T (1985) Alterations of regional cerebral blood flow and oxygen metabolism in Parkinson's disease. Neurology 35:1399–1405
- Wooten GF, Collins RC (1981) Metabolic effects of unilateral lesion of the substantia nigra. J Neurosci 1:285–291